

## GENETIC BASIS OF STRESS TOLERANCE IN RICE

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**Abstract:** Rice (*Oryza sativa L.*) is an essential diet for almost 50% of the global population. Rice harvests are vulnerable to a variety of living and non-living stresses. Pest insects, fungi, bacteria, viruses, and herbicide toxicity are a few examples of biotic stressors. Drought, cold, and salinity are three abiotic conditions that rice has also been extensively affected. Several genes have been discovered, cloned, and described to counteract these challenges and safeguard rice crops. Transgenic plants are created by successfully introducing the identified genes into rice plants. Rice crop improvement is significantly impacted by genetic engineering. This review article discusses the increased rice quality features tolerating living and non-living stress. This review's goal is to give readers a summary of recent advancements in rice biotechnology research and development.

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### Introduction

Rice was one of the first crops to be domesticated (*Oryza sativa L.*). For thousands of years, it has been raised in China and India. (Poehlman plus Sleper, 1995). *Oryza sativa* (2n = 24), Most widely farmed classes of rice, is a tropical Asian native. Only in western tropical Africa are the different rice classes grown, *Oryza glaberrima* (2n = 24), which remains native ascending the Niger River's upper valley. Ten identified genome varieties are AA, BB, CC, BBCC, CCDD, EE, FF, GG, HHJJ, and HHKK; 23 species; of Rice are known. The untamed *O. rufipogon*, a perennial plant, and the untamed *O. nivara*, a perennial are close cousins of *O. sativa*. Both weedy species are diploid and have the AA genome. Because of the ongoing population growth, rice must continually improve as a food crop. The following main goals are sought to enhance rice crops using biotechnological methods. Among the characteristics that these crops may have been high yield potential, early maturity, resistance to lodging and shattering, environmental stress tolerance, disease and pest resistance, increased grain quality, and improved nutritional components.

### The value of rice to the economy

Apart from Southeast Asia, China, India, Pakistan, Japan, and Korea nearby regions produce rice accounts for 90% of global production (USDA, 2014). The output from Brazil and the United States is most rice outside the Asia (1995; Sleper and Poehlman). Almost half of the globe's population relies mainly on

eating sources of rice (FAO, 2008). *Oryza* is the staple food crop that has a significant transfer from Pakistan. It contributes 0.7% to the GDP, and 3.2% to all agricultural value added. The expected area sown for rice is 2.891 106 ha, while the yield in 2014–2015 was 7.005 106 t. (Ministry of Finance, 2015). In Pakistan, rice is grown under a variety of weather circumstances. In Punjab Province's traditional rice-growing region, basmati rice (Indica) is farmed. Japanese rice grown in Swat is temperate and grows in high-altitude valleys. The majority of Long-grain tropical rice of the IRRI type is cultivated in the southern regions between Baluchistan and Sindh.

### Crop transformation in rice fields

Our Globe would 25 percent more rice is required by 2030 to fulfil the anticipated demand for a growing global people (Wani and Sah, 2014). Growing rice on more land is one approach to addressing this problem, although doing so is challenging given the rapid urbanisation and population growth in countries with low development. The alternative is to boost per-hectare production and improve varieties through breeding initiatives using traditional techniques and contemporary biotechnology. Biotechnology promises to increase crop productivity and reduce loss from diverse living and non-living challenges (Mahmood-ur-Rahman et al., 2014b; Ozawa and Takaiwa, 2010; Gelvin, 2010).

### Rice that resists insects

Modern agriculture has been transformed by insect-resistant crops, which are now a key component of integrated pest control strategies, reducing the need for insecticides while safeguarding environmental issues and Community health (Brooks and Barfoot, 2013). In the same way, other crops were created, insect-resistant rice plants were around for around 20 years (Fujimoto et al., 1993). Currently, field tests of genetically modified rice plants (Bt rice plants) that express a trait taken by Bt rice are being conducted in several nations (Wang et al. 2014; High et al. 2004; Tu et al. 2000). Several research suggest rice that is BT can diminish brought on via lepidopteran pests inside Asia (High et al., 2004). Using an artificial gene i.e. cry1Ab, transgenic rice turned out to be much more 8 Lepidoptera, as well as stripe-headed stem borer, resistant (*Chilo suppressalis*; SSB) plus Shu et al. state that stem borer in yellow (2000) (YSB; incertulas of the Scirpophaga). Also, dual strains of Bt *Oryza* plantations exhibited significant lepidopteran resistance. Pests in outdoor settings (Deka et al., 2010; Wang et al. 2014; Kumar et al. 2008). Table 1 reviews significant turning points in the emergence of rice plant life that remains insect-resistant.

At a field in China, insect-resistant hybrid rice Testing revealed that plants were extremely resistant to the YSB (Tu et al., 2000; Deka and Barthakur, 2010; Chen et al., 2011). The rice leaf folder (RLF; *Cnaphalocrocis medinalis*). Moreover, insect-resistant Bt rice has been developed in Pakistan and the Mediterranean. (Mahmood-ur-Rahman et al., 2007; Breitler et al., 2004). Current studies conducted in mutual areas revealed great resistance to the YSB and RLF, the target insects (Breitler et al. 2004; Bashir et al., 2005; Mahmood-ur-Rahman et al. 2007, 2012, 2013, 2014a, 2014b; Tabashnik et al., 2009) (Table 2). The target insects weren't naturally introduced to rice plants (Figures 1A–1C), and the intensity of their attack was quantified. India has also created certain transgenic rice lines and types resistant to YSB (Ramesh et al. 2004). In China, transgenic rice plants have been created in both fields and laboratories experiments have been conducted or done to see how effective they are (Li, 2013; Li et al. 2014; Wang et al. 2012). It particularly effective in creating long-lasting resistance against insect pests by pyramiding numerous genes against a certain pest or a variety of insects. Cry1Ab or Cry1Ac have been studied to pyramid with either.

**Table 1. Enhancing rice genetically to withstand insects**

genes	Aims	Reference
cry1Ab and cry1Ac	SSB, YSB, and SSB**	Su et al., (2000)
cry1Aa and cry1Ab		Breitler et al., (2004)
cry1Ab or cry1Ac	SSB	Ramesh et al., (2004)
cry1Ab	RLF and YSB ***	
cry1Ab	Sheath blight, bacterial blight, and YSB	Cotsaftis et al., (2002)
cry, Xa21, or RC7	Insects that are homopteran, coleopteran, and lepidopteran	Bashir et al., (2005)
gna or cry1Ac	Wheat weevil	Datta et al., (2003)
Itr1	RLF and YSB	Nagadhara et al., (2003)
cry1Ac or cry2A	Bug resistance	Alfonso-Rubi et al., (2003)
Bt or CpT1	Bug opposition	Mahmood-ur-Rahman et al., (2007)
Bacillus thuringiensis, lectins from plants, enzymes, or protease inhibitors	Bug opposition	Rong et al., (2007)
cry2Aa	Bug opposition	Deka and Barthakur, (2010)
cry1Ab	SSB, YSB, and SSB**	Wang et al., (2012)

**Table # 2. GM Rice evaluation in the Fields**

genes	Characteristics	Sites	Reference
cry1Ab or cry1Ac	resist insects	China	Tu et al., (2000)
cry1Ab	resist insects	China	Su et al., (2000)
cry1Ac or cry2A	resist insects	Pakistan	Bashir et al., (2005)
cry1Aa or cry1B		Spain	Breitler et al., (2004)
Cry2A	resist insects	China	Chen et al., (2005)
cry1Ac or cry2A	resist insects	Pakistan	(Mahmood-ur-Rahman et al., 2007, 2012, 2014a)s
cry1Ac or CpT1	Resist insects	China	Han et al., (2006)
cry1Ab	Resist insects	China	Wang et al., (2014)
Xa21	Resist insects	China	Tu et al., (2000)
bar	blight resistant to bacteria	United states of America	Oard et al., (2000)
bar	Drug-resistant weeds	Spain	Messeguer et al., (2004)
bar	Drug-resistant weeds	United states of America	Zhang et al., (2004)

For enhanced and long-lasting resistance in Bt rice, use cry2A or cry9C. Cry1Ab and Cry1Ac have the same binding site in insects like SSB and YSB and may be particularly successful at pyramiding genes (Alcantara et al., 2004). Increased lectin resistance for the variety was achieved by combining Cry genes with the snowdrop (*Galanthus nivalis*) lectin gene i.e insects, sucking included insects (Ramesh et al., 2004). Insect pests that suckers have no effect from the Bt genes (Bernal et al., 2002). To create a long-lasting resistance, it's crucial to stack additional insecticidal genes and cry genes. Plants may include proteins that prevent ribosomes from forming, lectins, or protease inhibitors that can be used to produce these genes for non-Bt insect resistance (Sharma et al., 2003). a gene for lectin (*Galanthus nivalis* agglutinin) has been extensively utilized to create rice with built-in resistance to insects of the Pest species belong to the homopteran, lepidopteran, and coleopteran orders (Nagadhara et al., 2003). Researchers have created and field-tested transgenic rice strains that generate proteinase inhibitors (Mochizuki et al 1999). A Commercial use using genetically engineered rice may benefit from recent advancements such a transgenic expression that is tissue-specific and inducible, pyramiding of many sets of genes greater spectrum delays in the emergence of insect pests' resistant strains and measures of protection (Kumar et al., 2008). Lectins and Gna (Lopez et al., 2002) is less effective than anti-protease agents (Hernandez et al., 2003). It may be possible to create insect-resistant rice using lectins and protease inhibitors that are more efficient against sucking insects (Kumar et al., 2008).

#### Fungus-Resistive Rice

Given that this yield is liable for the variety to infections, among them are viruses, bacteria, and fungus, genetic modification of fungal resistance of rice is urgently needed (Gust and others, 2010, Miah et al., 2013, Sattari et al., 2014). Many genes were found to provide resistance to several fungus species, identified, replicated, then transformed to rice plants. Recently, the Pi-ta gene was cloned, it was discovered to exhibit high resistance to rice disease i.e Blast (Delteil et al., 2010; Dai et al., 2010). Study and mapping of indigenous resistance to sheath blight (Liu et al., 2009). Also, genetic indicators were discovered through cross-breeding between susceptible/non-transgenic and transgenic types (Liu et al., 2009). Generations of Asian rice with the PR-3 rice chitinase gene were discovered to resist hermetic blight (Datta et al., 2003). The group of defense- similar Genes includes Rir1b Gene. It was just discovered. and described in the Grains (Mauch et al., 2000). Rir1b-containing Gene-modified Rice was more resistant to Rice Blast (Li et al. 2009). Also, numerous proteins were discovered to be helpful for candidates to give rice plants resistance against various fungal species, which help to facilitate pathogen assault. Examples include the selenium-binding protein homolog (Sawada & al., 2004), the puroindoline proteins

(Krishnamurthy et al., 2001), the lipid transfer protein (Guiderdoni et al., 2002), and the rice homolog of maize HC-toxin reductase (Uchimiya et al., 2001; Higa et al., 2003).

#### Bacteria-resistant rice

By modifying the endogenous gene Xa21, transgenic rice blight-resistant strains that resist germs were created (Song et al., 1995). Several Rice cultivars have Xa21 introduced through both genetic modification and traditional breeding methods ((Mohanty and Tyagi, 2000). There have been tests on Xa21-transgenic plants in the field with encouraging outcomes (Tu et al., 2000). The top option for creating bacterial blight resistance so far was discovered to be this gene. Additionally, it was stated that the gene and other genes could give several forms of stress tolerance (Datta et al., 2003). The rice Gene Xa 26, which expresses the same protein as similar effects in transgenic plants to those in Xa21-expressing plants, was also found to be resistant (Zhang, 2009; Sun et al., 2004). To create rice that is resistant to germs, certain additional genes have also been altered (Zhou et al., 2011). A class of the Genes called cecropins has antibacterial properties. In Cecropia moth hemolymph, they express peptides. They have transformed and have successfully expressed themselves in plants (Huang et al., 1997). AP1, a ferredoxin-like protein produced by transgenic rice plants, displayed ability to tolerate *X. oryzae* (Tang et al., 2001). Moreover, *Burkholderia plantarii* resistance has been induced in rice. Due to decreased pathogen damage, rice plants that express Xa21 in the field exhibit a considerable boost in production (Tu et al., 2000). For long-lasting resistance to bacterial infections in rice, the commercial release of the promising candidate gene Xa21 should benefit all farmers Kumar et al. (2008)

#### Virus-resistant rice

For many years, yield loss brought on by viruses has been a major issue worldwide. In particular, the rice dwarf virus (RDV) and the rice stripe virus (RSV), which led to significant production losses during 1960 (2010) (Toriyama), are now more likely to cause damage. Insecticides can be rice fields may be managed using the method used to control vector insects, however their high cost and environmental risk are the main barriers. One of the best ways to shield rice plants against viral infection is through the evolution of rice virus resistance genetically or the insects that serve as their insect vectors. Researchers created and assessed rice plants resistant to RSV and RDV (Sasaya et al., 2013; Shimizu et al., 2009; Xiong et al., 2009). The viral rice illness known as Rice tungro is quite dangerous. RTBV (rice tungro bacilliform virus) and RTSV (rice tungro spherical virus) are two viruses. —are responsible for its development (RTSV). The green leafhopper (*Nephotettix virescens*) acts as RTSV's vector and aids in the spread of the disease. Using the coat, rice plants have changed protein-mediated defence mechanisms

(Huet et al. in 1999). Developed transgenic rice plants with the RTSV replicase gene. The transgenic plants demonstrated a minor amount of RTSV resistance.

#### Rice resistance to herbicide

Agrochemical resistance is a crucial agronomic feature that has been effectively employed for many years to eliminate weeds. The EPSPS and bar genes are only two of the many genes being exploited to create herbicide-tolerant plants. The EPSPS gene was acquired from *Agrobacterium* strain CP4 and detoxifies glyphosate herbicides, while the bar gene was taken from *Streptomyces hygroscopicus*, which detoxifies the herbicide glufosinate (Kumar et al., 2008). In the early days of rice transformation research, herbicide-resistant GM rice plants carrying the bar gene were created; these plants are currently undergoing field testing (Tyagi and Mohanty, 2000; Oard et al., 2000). It has taken a lot of labour to find other gene sources that could be exploited to give transgenic rice long-lasting resistance to herbicides. One such instance is the protoporphyrinogen oxidase produced by the bacterium *Bacillus subtilis*, which rice plants effectively use to develop resistance to the oxyfluorfen herbicide (Jung et al., 2004). Another illustration is people's Cytochrome P450s, which, when overexpressed in rice led to inconsistent herbicide reactions (Kawahigashi and co., 2003).

#### Rice Resistant to abiotic stress

Droughts, extreme heat or cold, salinity, submersion, and oxidative stress are abiotic factors that severely lower crop productivity. These pressures have been documented to cause crop loss of more than 50% internationally (Iqbal et al. 2013; Li et al., 2016; Bray et al., 2000). It frequently interacts with one another

and result in similar cellular and physiological harm. Moreover, they stimulate similar mechanisms for cell signaling (Qin et al., 2011); Nakashima et al. (2009). Several proteins, antioxidants, and suitable solutes are created in response to stressful situations. Many agricultural plants have been created by overexpressing the genes for these chemicals, and they have been tested in both the lab and the field for many different abiotic stresses (Luo et al. 2010). The worst elements that reduce rice crop output are tolerance to water scarcity and salt stress. One method to improve crop plants' ability to withstand abiotic stress is GM technology (Flowers, 2004). Creating transgenic plants with various genes that produce drought and/or salinity tolerance in multiple plants has helped crop species like rice (Tyagi and Mohanty, 2000; Iqbal et al., 2013). (Table 3). Moreover, many new genes have been discovered and identified in model plants that confer tolerance to salt and drought stress (reviewed by Flowers, 2004; Zhang et al., 2004).

Using the chloroplastic glutamine synthase (GS2) gene, Hoshida et al. (2000) successfully created transgenic rice plants. Strong salt and freezing stress tolerance and improved photorespiration abilities have been demonstrated in transgenic plants with high GS2 levels (Hoshida et al., 2000). OsMAPK5, a stress-sensitive MAPK gene that responds to ABA and other biotic and abiotic stimuli in rice, was found and described by Xiong and Yang in 2003. OsMAPK5 overexpression makes plants more resistant to salt, cold, and drought stress (Xiong and Yang, 2003; Osakabe et al., 2014; Savchenko et al., 2014)

**Table 3. *Oryza sativa* genes that can withstand abiotic stress**

S. No.	genes	Purpose	Origin	Features of transgenic plants	reference
1.	GS2	glucose synthesis	<i>Oryza sativa</i>	tolerance of the effects of cold and salt	Hoshida et al., (2000)
2.	OsCDPK7	PTKs that is calcium-dependent	Rice	tolerant of the stresses of drought and salinity	Saijo et al., (2000)
3.	OsMAPK5	A Mitogen-activated protein	<i>Oryza sativa</i>		Yang and Xiong (2003)
4.	Adc, Samdc		Oats		
5.	HVA1	Late embryogenesis abundant proteins polyamine biosynthesis	Barley and Wheat	tolerance of the stresses of drought and salinity	Babu et al. (2004); Capell et al. (2004)
6.	OtsA	Anabolism of trehalose	Bacteria	tolerance of the stresses of dehydration and salt	Jung et al., (2003)
7.	p5cs	prostaglandin synthesis	dew gram	Salt, drought, and cold stress resistance	Hur et al., (2004); Rahman et al (2001)

8.	pdc1, adc	(ODC) of pyruvate	Oryza sativa	Transgene expression response to stress	Ariizumi (2002)	et al.,
9.	AGPAT, SGPAT	Formulation of lipids	Spinacia oleracea	tolerance for submersion	Matsumura (2003); Yamanouchi et al (2002)	
10.	Cat	Catalase	Triticum	enhanced photosynthetic efficiency at cold temperatures	Hoshida et al., (2000)	
11.	spl7	heat stress transcription factor	Oryza sativa	Cold-tolerant	Saijo et al., (2000)	

In response to abiotic stress, dehydration-responsive elements (DREs) control the expression of numerous genes (Nakashima et al., 2014; Wani and Sah, 2014). OsDREB1A, OsDREB1B, OsDREB1C, OsDREB1D, and OsDREB2A are the five rice DREB homologs of *Arabidopsis* that were discovered and characterized by Dubouzet et al (2003). Several OsDREB genes were overexpressed in transgenic rice plants while regulated by various promoter combinations. They asserted that OsDREB1A was especially useful in generating transgenic dicot and monocot plants that were more resistant to the impacts of cold, salt, and/or drought (Nawaz et al., 2014; Wang et al., 2007).

The ACS gene causes rice plants' resistance to submersion. The vascular bundles of juvenile stems and leaf sheaths showed greater mRNA levels when the plant was partially immersed. The same outcomes were also seen in rice when the YK1 gene was overexpressed (Uchimiya et al., 2002). Spl7, a transcription factor, was identified and cloned in rice plants. In spl7 mutants, its overexpression reduced the development of leaf spots brought on by high temperatures (Yamanouchi et al., 2002). Furthermore, heat shock proteins or oxidative stress-tolerance enzymes have been overexpressed in rice plants to produce heat-tolerant rice plants (Murakami et al., 2004; Kouril et al., 2003)

### Conclusion

Regarding global food security and sustainable agricultural methods, the genetic underpinnings of rice's ability to withstand biotic and abiotic stress are a required field of study. One of the major food crops on the planet, rice gives millions of people a significant source of calories and nourishment. Unfortunately, rice crops are vulnerable to several biotic and abiotic stressors, which can seriously reduce productivity and jeopardize food security. Because of the importance of rice economically and the difficulties brought on by biotic and abiotic stressors, creative solutions are needed. The production of rice cultivars with improved stress tolerance is made possible through genetic engineering, which offers a viable strategy for

overcoming these difficulties. The production of rice variants with increased stress tolerance has been made possible through the genetic engineering of rice crops. A few examples of the genetic alterations created to address the problems facing the rice business are insect resistance rice, fungus resistance rice, bacterium resistance rice, virus resistance rice, and herbicide tolerance rice. These improvements may boost crop yields and enhance food security in areas where rice is a staple crop. The transmission of stress-induced alterations also complicates the genetic foundation for stress tolerance in rice through generations and our emerging understanding of epigenetic modifications. This opens up new opportunities for creating rice types more resistant to stress. To make sure that the potential advantages of genetic engineering have been achieved while avoiding the hazards. It is necessary to approach genetic engineering research cautiously, focusing on safety evaluations and regulatory frameworks. It is crucial to weigh the possible advantages against the potential drawbacks to make new genetic tools and technologies safe for the environment and people's health. Therefore, it is essential to continue studying the genetic underpinnings of rice's ability to withstand biotic and abiotic stress to promote sustainable agricultural methods, ensure global food security, and solve issues affecting the rice sector. The global rice business might change if more robust, high-yielding, and nutrient-dense rice varieties are created, allowing farmers to produce more food with fewer resources and maintaining food security for millions of people.

### Conflict of interest

The authors declared absence of conflict of interest.

### References

Alcantara, E. P., Aguda, R. M., Curtiss, A., Dean, D. H., & Cohen, M. B. (2004). *Bacillus thuringiensis*  $\delta$ -endotoxin binding to brush border membrane vesicles of rice stem borers. *Archives of Insect Biochemistry and Physiology: Published in Collaboration with the Entomological Society of America*, 55(4), 169-177.

Alfonso-Rubí, J., Ortego, F., Castañera, P., Carbonero, P., & Díaz, I. (2003). Transgenic expression of trypsin inhibitor CMe from barley in indica and japonica rice, confers resistance to the rice weevil *Sitophilus oryzae*. *Transgenic Research*, **12**, 23-31.

Ansari, M. U. R., Shaheen, T., Bukhari, S., & Husnain, T. (2015). Genetic improvement of rice for biotic and abiotic stress tolerance. *Turkish Journal of Botany*, **39**(6), 911-919.

Ariizumi, T., Kishitani, S., Inatsugi, R., Nishida, I., Murata, N., & Toriyama, K. (2002). An increase in unsaturation of fatty acids in phosphatidylglycerol from leaves improves the rates of photosynthesis and growth at low temperatures in transgenic rice seedlings. *Plant and Cell Physiology*, **43**(7), 751-758.

Babu, R. C., Zhang, J., Blum, A., Ho, T. H. D., Wu, R., & Nguyen, H. T. (2004). HVA1, a LEA gene from barley confers dehydration tolerance in transgenic rice (*Oryza sativa* L.) via cell membrane protection. *Plant Science*, **166**(4), 855-862.

Bashir, K., Husnain, T., Fatima, T., Riaz, N., Makhdoom, R., & Riazuddin, S. (2005). Novel indica basmati line (B-370) expressing two unrelated genes of *Bacillus thuringiensis* is highly resistant to two lepidopteran insects in the field. *Crop protection*, **24**(10), 870-879.

Bernal, C. C., Aguda, R. M., & Cohen, M. B. (2002). Effect of rice lines transformed with *Bacillus thuringiensis* toxin genes on the brown planthopper and its predator *Cyrtorhinus lividipennis*. *Entomologia Experimentalis et Applicata*, **102**(1), 21-28.

Bray, E. A. (2000). Responses to abiotic stresses. *Biochemistry and Molecular Biology of Plants*, 1158-1203.

Breitler, J. C., Vassal, J. M., Del Mar Catala, M., Meynard, D., Marfà, V., Melé, E., ... & Messeguer, J. (2004). Bt rice harbouring cry genes controlled by a constitutive or wound-inducible promoter: protection and transgene expression under Mediterranean field conditions. *Plant Biotechnology Journal*, **2**(5), 417-430.

Brookes, G., & Barfoot, P. (2013). Key environmental impacts of global genetically modified (GM) crop use 1996–2011. *GM Crops & Food*, **4**(2), 109-119.

Capell, T., Bassie, L., & Christou, P. (2004). Modulation of the polyamine biosynthetic pathway in transgenic rice confers tolerance to drought stress. *Proceedings of the National Academy of Sciences*, **101**(26), 9909-9914.

Chandio, A. A., Yuansheng, J., & Magsi, H. (2016). Agricultural sub-sectors performance: an analysis of sector-wise share in agriculture GDP of Pakistan. *International Journal of Economics and Finance*, **8**(2), 156-162.

Chen, H., Tang, W., Xu, C., Li, X., Lin, Y., & Zhang, Q. (2005). Transgenic indica rice plants harboring a synthetic cry2A\* gene of *Bacillus thuringiensis* exhibit enhanced resistance against lepidopteran rice pests. *Theoretical and Applied Genetics*, **111**, 1330-1337.

Chen, M., Shelton, A., & Ye, G. Y. (2011). Insect-resistant genetically modified rice in China: from research to commercialization. *Annual Review of Entomology*, **56**, 81-101.

Coca, M., Bortolotti, C., Rufat, M., Penas, G., Eritja, R., Tharreau, D., ... & San Segundo, B. (2004). Transgenic rice plants expressing the antifungal AFP protein from *Aspergillus giganteus* show enhanced resistance to the rice blast fungus *Magnaporthe grisea*. *Plant Molecular Biology*, **54**, 245-259.

Cotsaftis, O., Sallaud, C., Breitler, J. C., Meynard, D., Greco, R., Pereira, A., & Guideroni, E. (2002). Transposon-mediated generation of T-DNA-and marker-free rice plants expressing a Bt endotoxin gene. *Molecular Breeding*, **10**, 165-180.

Dai, Y., Jia, Y., Correll, J., Wang, X., & Wang, Y. (2010). Diversification and evolution of the avirulence gene AVR-Pita1 in field isolates of *Magnaporthe oryzae*. *Fungal Genetics and Biology*, **47**(12), 973-980.

Datta, K., Baisakh, N., Oliva, N., Torrizo, L., Abrigo, E., Tan, J., ... & Datta, S. K. (2003). Bioengineered 'golden' indica rice cultivars with β-carotene metabolism in the endosperm with hygromycin and mannose selection systems. *Plant Biotechnology Journal*, **1**(2), 81-90.

Deka, S., & Barthakur, S. (2010). Overview on current status of biotechnological interventions on yellow stem borer *Scirpophaga incertulas* (Lepidoptera: Crambidae) resistance in rice. *Biotechnology advances*, **28**(1), 70-81.

Delteil, A., Zhang, J., Lessard, P., & Morel, J. B. (2010). Potential candidate genes for improving rice disease resistance. *Rice*, **3**(1), 56-71.

Dubouzet, J. G., Sakuma, Y., Ito, Y., Kasuga, M., Dubouzet, E. G., Miura, S., ... & Yamaguchi-Shinozaki, K. (2003). OsDREB genes in rice, *Oryza sativa* L., encode transcription activators that function in drought-, high-salt-and cold-responsive gene expression. *The Plant Journal*, **33**(4), 751-763.

Flowers, T. J. (2004). Improving crop salt tolerance. *Journal of Experimental Botany*, **55**(396), 307-319.

Fujimoto, H., Itoh, K., Yamamoto, M., Kyozuka, J., & Shimamoto, K. O. (1993). Insect resistant rice generated by introduction of a modified δ-endotoxin gene of *Bacillus thuringiensis*. *Bio/technology*, **11**(10), 1151-1155.

Gandikota, M., Kochko, A. D., Chen, L., Ithal, N., Fauquet, C., & Reddy, A. R. (2001). Development of transgenic rice plants

expressing maize anthocyanin genes and increased blast resistance. *Molecular Breeding*, **7**(1), 73-83.

Garnett, T., Appleby, M. C., Balmford, A., Bateman, I. J., Benton, T. G., Bloomer, P., ... & Godfray, H. C. J. (2013). Sustainable intensification in agriculture: premises and policies. *Science*, **341**(6141), 33-34.

Gelvin SB (2010). Plant proteins involved in Agrobacterium-mediated genetic transformation. *Annu Rev Phytopathol* **48**: 45–68.

Guiderdoni, E., Cordero, M. J., Vignols, F., Garcia-Garrido, J. M., Lescot, M., Tharreau, D., ... & Delseny, M. (2002). Inducibility by pathogen attack and developmental regulation of the rice Ltp1 gene. *Plant Molecular Biology*, **49**(6), 679-695.

Gust, A. A., Brunner, F., & Nürnberg, T. (2010). Biotechnological concepts for improving plant innate immunity. *Current Opinion in Biotechnology*, **21**(2), 204-210.

Han, L. Z., Wu, K. M., Peng, Y. F., Wang, F., & Guo, Y. Y. (2006). Evaluation of transgenic rice expressing Cry1Ac and CpTI against Chilo suppressalis and intrapopulation variation in susceptibility to Cry1Ac. *Environmental entomology*, **35**(5), 1453-1459.

Handberg, R. (1993). Space Policy: An Introduction-Nathan C. Goldman, Ames, Iowa: Iowa State University Press, 1992, 332 pp. ISBN 0-8138-1024-8. Iowa State University Press, 2121 S. State Ave., Ames, IA 50010, USA. *Politics and the Life Sciences*, **12**(2), 305-306.

Hernández, C. A., Pujol, M., Alfonso-Rubí, J., Armas, R., Coll, Y., Pérez, M., ... & Ortego, F. (2003). Proteolytic gut activities in the rice water weevil, *Lissorhoptrus brevirostris* Suffrian (Coleoptera: Curculionidae). *Archives of Insect Biochemistry and Physiology: Published in Collaboration with the Entomological Society of America*, **53**(1), 19-29.

Higa, A., Kimura, M., Mimori, K., Ochiai-Fukuda, T., Tokai, T., Takahashi-Ando, N., ... & Yamaguchi, I. (2003). Expression in cereal plants of genes that inactivate Fusarium mycotoxins. *Bioscience, biotechnology, and Biochemistry*, **67**(4), 914-918.

High, S. M., Cohen, M. B., Shu, Q. Y., & Altosaar, I. (2004). Achieving successful deployment of Bt rice. *Trends in Plant Science*, **9**(6), 286-292.

Hoshida, H., Tanaka, Y., Hibino, T., Hayashi, Y., Tanaka, A., Takabe, T., & Takabe, T. (2000). Enhanced tolerance to salt stress in transgenic rice that overexpresses chloroplast glutamine synthetase. *Plant Molecular Biology*, **43**(1), 103.

Huang, N., Angeles, E. R., Domingo, J., Magpantay, G., Singh, S., Zhang, G., ... & Khush, G. S. (1997). Pyramiding of bacterial blight resistance genes in rice: marker-assisted selection using RFLP and PCR. *Theoretical and Applied Genetics*, **95**, 313-320.

Huet, H., Mahendra, S., Wang, J., Sivamani, E., Ong, C. A., Chen, L., ... & Fauquet, C. (1999). Near immunity to rice tungro spherical virus achieved in rice by a replicase-mediated resistance strategy. *Phytopathology*, **89**(11), 1022-1027.

Hur, J., Jung, K. H., Lee, C. H., & An, G. (2004). Stress-inducible OsP5CS2 gene is essential for salt and cold tolerance in rice. *Plant Science*, **167**(3), 417-426.

Iqbal, M., Khan, M. A., Naeem, M., Aziz, U., Afzal, J., & Latif, M. (2013). Inducing drought tolerance in upland cotton (*Gossypium hirsutum* L.), accomplishments and future prospects. *World Appl. Sci. J.*, **21**(7), 1062-1069.

Jung, S., Chung, J. S., Chon, S. U., Kuk, Y. I., Lee, H. J., Guh, J. O., & Back, K. (2004). Expression of recombinant protoporphyrinogen oxidase influences growth and morphological characteristics in transgenic rice. *Plant growth regulation*, **42**, 283-288.

Kanzaki, H., Nirasawa, S., Saitoh, H., Ito, M., Nishihara, M., Terauchi, R., & Nakamura, I. (2002). Overexpression of the wasabi defensin gene confers enhanced resistance to blast fungus (*Magnaporthe grisea*) in transgenic rice. *Theoretical and Applied Genetics*, **105**(6-7), 809.

Kawahigashi, H., Hirose, S., Ohkawa, H., & Ohkawa, Y. (2003). Transgenic rice plants expressing human CYP1A1 exude herbicide metabolites from their roots. *Plant Science*, **165**(2), 373-381.

Kouřil, R., Lazar, D., Lee, H., Jo, J., & Nauš, J. (2003). Moderately elevated temperature eliminates resistance of rice plants with enhanced expression of glutathione reductase to intensive photooxidative stress. *Photosynthetica*, **41**, 571-578.

Krishnamurthy, K., Balconi, C., Sherwood, J. E., & Giroux, M. J. (2001). Wheat puroindolines enhance fungal disease resistance in transgenic rice. *Molecular Plant-Microbe Interactions*, **14**(10), 1255-1260.

Kumar, S., Chandra, A., & Pandey, K. C. (2008). *Bacillus thuringiensis* (Bt) transgenic crop: an environment friendly insect-pest management strategy. *J Environ Biol*, **29**(5), 641-653.

Lee, H. J., Lee, D. E., Ha, S. B., Jang, S. W., Lee, I. J., Ryu, S. B., & Back, K. (2004). The characterization of transgenic rice plants expressing a pepper 5-epi aristolochene synthase, the first committed step enzyme for capsidiol synthesis in isoprenoid pathway. *Plant science*, **166**(4), 881-887.

Li, F., Fan, G., Wang, K., Sun, F., Yuan, Y., Song, G., ... & Yu, S. (2014). Genome sequence of the cultivated cotton *Gossypium arboreum*. *Nature genetics*, **46**(6), 567-572.

Li, H., Zhou, S. Y., Zhao, W. S., Su, S. C., & Peng, Y. L. (2009). A novel wall-associated receptor-like protein kinase gene, OsWAK1, plays important roles in rice blast disease resistance. *Plant molecular Biology*, **69**, 337-346.

Li, Y., Hu, L., Romeis, J., Wang, Y., Han, L., Chen, X., & Peng, Y. (2014). Use of an artificial diet system to study the toxicity of gut-active insecticidal compounds on larvae of the green lacewing Chrysoperla sinica. *Biological Control*, **69**, 45-51.

Li, Y., Wang, Y., Romeis, J., Liu, Q., Lin, K., Chen, X., & Peng, Y. (2013). Bt rice expressing Cry2Aa does not cause direct detrimental effects on larvae of Chrysoperla sinica. *Ecotoxicology*, **22**, 1413-1421.

Liu, G. U. A. N. G. J. I. E., Jia, Y., Correa-Victoria, F. J., Prado, G. A., Yeater, K. M., McClung, A., & Correll, J. C. (2009). Mapping quantitative trait loci responsible for resistance to sheath blight in rice. *Phytopathology*, **99**(9), 1078-1084.

Lopez, S., Codina, C., Bastida, J., Viladomat, F., Davidson, E., & Stewart, D. (2002). Biodiversity of mannose-specific lectins within Narcissus species. *Journal of Agricultural and Food Chemistry*, **50**(9), 2507-2513.

Luo, M., Liu, J., Lee, R. D., Scully, B. T., & Guo, B. (2010). Monitoring the expression of maize genes in developing kernels under drought stress using oligo-microarray. *Journal of Integrative Plant Biology*, **52**(12), 1059-1074.

Mahmood-ur-Rahman, N. M., Shahid, A., Rao, A., & Husnain, T. (2014). Field performance and biosafety studies of segregating generations of transgenic basmati rice (*Oryza sativa* L.) containing multiple genes from *Bacillus thuringiensis*. *Annals of Agriculture Science (Special Issue)*, **1**, 39-42.

Mahmood-ur-Rahman, R. A., Batool, F., Azam, S., Shahid, A. A., & Husnain, T. (2012). Transgene copy number and phenotypic variations in transgenic basmati rice. *J Anim Plant Sci*, **22**, 1004-1013.

Matsumura, H., Nirasawa, S., Kiba, A., Urasaki, N., Saitoh, H., Ito, M., ... & Terauchi, R. (2003). Overexpression of Bax inhibitor suppresses the fungal elicitor-induced cell death in rice (*Oryza sativa* L.) cells. *The Plant Journal*, **33**(3), 425-434.

Mauch, F., Reimann, C., Freydl, E., Schaffrath, U., & Dudler, R. (1998). Characterization of the rice pathogen-related protein Rir1a and regulation of the corresponding gene. *Plant molecular biology*, **38**, 577-586.

Miah, G., Rafii, M. Y., Ismail, M. R., Puteh, A. B., Rahim, H. A., Asfaliza, R., & Latif, M. A. (2013). Blast resistance in rice: a review of conventional breeding to molecular approaches. *Molecular Biology Reports*, **40**, 2369-2388.

Mochizuki, A., Nishizawa, Y., Onodera, H., Tabei, Y., Toki, S., Habu, Y., ... & Ohashi, Y. (1999). Transgenic rice plants expressing a trypsin inhibitor are resistant against rice stem borers, *Chilo suppressalis*. *Entomologia Experimentalis et Applicata*, **93**(2), 173-178.

Murakami, T., Matsuba, S., Funatsuki, H., Kawaguchi, K., Saruyama, H., Tanida, M., & Sato, Y. (2004). Over-expression of a small heat shock protein, sHSP17. 7, confers both heat tolerance and UV-B resistance to rice plants. *Molecular Breeding*, **13**, 165-175.

Nagadhara, D., Ramesh, S., Pasalu, I. C., Rao, Y. K., Krishnaiah, N. V., Sarma, N. P., ... & Rao, K. V. (2003). Transgenic indica rice resistant to sap-sucking insects. *Plant Biotechnology Journal*, **1**(3), 231-240.

Nakashima, K., Ito, Y., & Yamaguchi-Shinozaki, K. (2009). Transcriptional regulatory networks in response to abiotic stresses in *Arabidopsis* and grasses. *Plant physiology*, **149**(1), 88-95.

Nawaz, M., Iqbal, N., Idrees, S., & Ullah, I. (2014). DREB1A from *Oryza sativa* var. IR6: homology modelling and molecular docking. *Turkish Journal of Botany*, **38**(6), 1095-1102.

Oard, J., Cohn, M. A., Linscombe, S., Gealy, D., & Gravos, K. (2000). Field evaluation of seed production, shattering, and dormancy in hybrid populations of transgenic rice (*Oryza sativa*) and the weed, red rice (*Oryza sativa*). *Plant Science*, **157**(1), 13-22.

Osakabe, Y., Osakabe, K., Shinozaki, K., & Tran, L. S. P. (2014). Response of plants to water stress. *Frontiers in Plant Science*, **5**, 86.

Ozawa, K., & Takaiwa, F. (2010). Highly efficient Agrobacterium-mediated transformation of suspension-cultured cell clusters of rice (*Oryza sativa* L.). *Plant Science*, **179**(4), 333-337.

Qasim, M., Bukhari, S. A., & Shaheen, T. (2014). Bt crops: a sustainable approach towards biotic stress tolerance. In *Emerging Technologies and Management of Crop Stress Tolerance* (pp. 125-142). Academic Press.

Qin, F., Shinozaki, K., & Yamaguchi-Shinozaki, K. (2011). Achievements and challenges in understanding plant abiotic stress responses and tolerance. *Plant and Cell Physiology*, **52**(9), 1569-1582.

Qu, L. J., Chen, J., Liu, M., Pan, N., Okamoto, H., Lin, Z., ... & Chen, Z. (2003). Molecular cloning and functional analysis of a novel type of Bowman-Birk inhibitor gene family in rice. *Plant physiology*, **133**(2), 560-570.

Rahman, M. U., Rashid, H., Shahid, A. A., Bashir, K., Husnain, T., & Riazuddin, S. (2007). Insect resistance and risk assessment studies of advanced generations of basmati rice expressing two genes of *Bacillus thuringiensis*. *Electronic Journal of Biotechnology*, **10**(2), 241-251.

Rahman, M., Grover, A., Peacock, W. J., Dennis, E. S., & Ellis, M. H. (2001). Effects of manipulation of pyruvate decarboxylase and alcohol dehydrogenase levels on the submergence tolerance of rice. *Functional Plant Biology*, **28**(12), 1231-1241.

Ramesh, S., Nagadhara, D., Reddy, V. D., & Rao, K. V. (2004). Production of transgenic indica rice resistant to yellow stem borer and sap-sucking insects, using super-binary vectors of *Agrobacterium tumefaciens*. *Plant Science*, **166**(4), 1077-1085.

Rong, J., Lu, B. R., Song, Z., Su, J., Snow, A. A., Zhang, X., ... & Wang, F. (2007). Dramatic reduction of crop-to-crop gene flow within a short distance from transgenic rice fields. *New Phytologist*, **173**(2), 346-353.

Saijo, Y., Hata, S., Kyozuka, J., Shimamoto, K., & Izui, K. (2000). Over-expression of a single  $\text{Ca}^{2+}$ -dependent protein kinase confers both cold and salt/drought tolerance on rice plants. *The Plant Journal*, **23**(3), 319-327.

Sasaya, T., Aoki, H., Omura, T., Yatou, O., & Saito, K. (2013). Development of virus-resistant transgenic forage rice cultivars. *Front. Microbiol*, **4**(409), 1997-2.

Sattari, A., Fakheri, B., Hassan, F. S. C., & Noroozi, M. (2014). Blast resistance in rice: a review of breeding and biotechnology. *International Journal of Agriculture and Crop Sciences (IJACS)*, **7**(6), 329-333.

Savchenko, T., Kolla, V. A., Wang, C. Q., Nasafi, Z., Hicks, D. R., Phadungchob, B., ... & Dehesh, K. (2014). Functional convergence of oxylipin and abscisic acid pathways controls stomatal closure in response to drought. *Plant Physiology*, **164**(3), 1151-1160.

Sawada, K., Hasegawa, M., Tokuda, L., Kameyama, J., Kodama, O., Kohchi, T., ... & Shinmyo, A. (2004). Enhanced resistance to blast fungus and bacterial blight in transgenic rice constitutively expressing OsSBP, a rice homologue of mammalian selenium-binding proteins. *Bioscience, Biotechnology, and Biochemistry*, **68**(4), 873-880.

Sharma, M., Charak, K. S., & Ramanaiah, T. V. (2003). Agricultural biotechnology research in India: Status and policies. *Current Science*, **84**(3), 297-302.

Shimizu, T., Yoshii, M., Wei, T., Hirochika, H., & Omura, T. (2009). Silencing by RNAi of the gene for Pns12, a viroplasm matrix protein of Rice dwarf virus, results in strong resistance of transgenic rice plants to the virus. *Plant biotechnology journal*, **7**(1), 24-32.

Shu, Q., Ye, G., Cui, H., Cheng, X., Xiang, Y., Wu, D., ... & Altosaar, I. (2000). Transgenic rice plants with a synthetic cry1Ab gene from *Bacillus thuringiensis* were highly resistant to eight lepidopteran rice pest species. *Molecular Breeding*, **6**, 433-439.

Sleper, D. A., & Shannon, J. G. (2003). Role of public and private soybean breeding programs in the development of soybean varieties using biotechnology. *AgBioForum*, **6**(1&2): 27-32.

Song, W. Y., Wang, G. L., Chen, L. L., Kim, H. S., Pi, L. Y., Holsten, T., ... & Ronald, P. (1995). A receptor kinase-like protein encoded by the rice disease resistance gene, Xa21. *Science*, **270**(5243), 1804-1806.

Strong, G. C. C. (2019). Grain: World markets and trade. <https://www.fas.usda.gov/data/grain-world-markets-and-trade>

Sun, X., Cao, Y., Yang, Z., Xu, C., Li, X., Wang, S., & Zhang, Q. (2004). Xa26, a gene conferring resistance to *Xanthomonas oryzae* pv. *oryzae* in rice, encodes an LRR receptor kinase-like protein. *The Plant Journal*, **37**(4), 517-527.

Tabashnik, B. E., Van Rensburg, J. B. J., & Carrière, Y. (2009). Field-evolved insect resistance to Bt crops: definition, theory, and data. *Journal of Economic Entomology*, **102**(6), 2011-2025.

Tang, K., Sun, X., Hu, Q., Wu, A., Lin, C. H., Lin, H. J., ... & Feng, T. (2001). Transgenic rice plants expressing the ferredoxin-like protein (AP1) from sweet pepper show enhanced resistance to *Xanthomonas oryzae* pv. *oryzae*. *Plant Science*, **160**(5), 1035-1042.

Toriyama, S. (2010). Rice Stripe: A Serious Threat to Rice Crops of Japan.

Tu, J., Datta, K., Khush, G. S., Zhang, Q., & Datta, S. K. (2000). Field performance of Xa21 transgenic indica rice (*Oryza sativa* L.), IR72. *Theoretical and Applied Genetics*, **101**, 15-20.

Tu, J., Zhang, G., Datta, K., Xu, C., He, Y., Zhang, Q., ... & Datta, S. K. (2000). Field performance of transgenic elite commercial hybrid rice expressing *Bacillus thuringiensis*  $\delta$ -endotoxin. *Nature biotechnology*, **18**(10), 1101-1104.

Tyagi, A. K., & Mohanty, A. (2000). Rice transformation for crop improvement and functional genomics. *Plant Science*, **158**(1-2), 1-18.

Uchimiya, H., Fujii, S., Huang, J., Fushimi, T., Nishioka, M., Kim, K. M., ... & Tagawa, M. (2002). Transgenic rice plants conferring increased tolerance to rice blast and multiple environmental stresses. *Molecular Breeding*, **9**, 25-31.

Wang, H., Zhang, H., Gao, F., Li, J., & Li, Z. (2007). Comparison of gene expression between upland and lowland rice cultivars under water stress using cDNA microarray. *Theoretical and Applied Genetics*, **115**, 1109-1126.

Wang, Y., Li, Y., Romeis, J., Chen, X., Zhang, J., Chen, H., & Peng, Y. (2012). Consumption of Bt rice pollen expressing Cry2Aa does not cause adverse effects on adult *Chrysoperla sinica*

Tjeder (Neuroptera: Chrysopidae). *Biological Control*, **61**(3), 246-251.

Wang, Y., Zhang, L., Li, Y., Liu, Y., Han, L., Zhu, Z., ... & Peng, Y. (2014). Expression of Cry1Ab protein in a marker-free transgenic Bt rice line and its efficacy in controlling a target pest, *Chilo suppressalis* (Lepidoptera: Crambidae). *Environmental Entomology*, **43**(2), 528-536.

Wani, S. H., & Sah, S. K. (2014). Biotechnology and abiotic stress tolerance in rice. *J Rice Res*, **2**(2), e105.

Xiong, L., & Yang, Y. (2003). Disease resistance and abiotic stress tolerance in rice are inversely modulated by an abscisic acid-inducible mitogen-activated protein kinase. *The Plant Cell*, **15**(3), 745-759.

Xiong, R., Wu, J., Zhou, Y., & Zhou, X. (2009). Characterization and subcellular localization of an RNA silencing suppressor encoded by Rice stripe tenuivirus. *Virology*, **387**(1), 29-40.

Yamanouchi, U., Yano, M., Lin, H., Ashikari, M., & Yamada, K. (2002). A rice spotted leaf gene, *Spl7*, encodes a heat stress transcription factor protein. *Proceedings of the National Academy of Sciences*, **99**(11), 7530-7535.

Yuan, H., Ming, X., Wang, L., Hu, P., An, C., & Chen, Z. (2002). Expression of a gene encoding trichosanthin in transgenic rice plants enhances resistance to fungus blast disease. *Plant cell reports*, **20**, 992-998.

Zhang, J. Z., Creelman, R. A., & Zhu, J. K. (2004). From laboratory to field. Using information from *Arabidopsis* to engineer salt, cold, and drought tolerance in crops. *Plant physiology*, **135**(2), 615-621.

Zhang, Q. (2009). Genetics and improvement of bacterial blight resistance of hybrid rice in China. *Rice Science*, **16**(2), 83-92.

Zhou, Y. L., Uzokwe, V. N., Zhang, C. H., Cheng, L. R., Wang, L., Chen, K., ... & Li, Z. K. (2011). Improvement of bacterial blight resistance of hybrid rice in China using the *Xa23* gene derived from wild rice (*Oryza rufipogon*). *Crop Protection*, **30**(6), 637-644.



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